

Molecular Dynamics Flexible Fitting

NAMD MDFF

<https://www.ks.uiuc.edu/Research/mdff/>

VMD

<https://www.ks.uiuc.edu/Research/vmd/>

Cryo-EM Map

Atomic Coordinates – PDB File, Homology model,
<https://www.rcsb.org/>

Molecular Mechanics Force Field

CHARMM36 - http://mackerell.umaryland.edu/charmm_ff.shtml

Amber - <http://ambermd.org/AmberModels.php>

$$U_{\text{Total}} = U_{\text{bonded}} + U_{\text{nonbonded}}$$

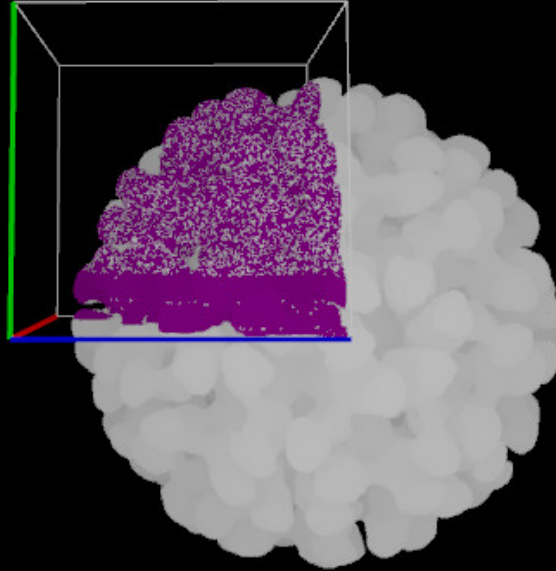
$$U_{\text{bonded}} = U_{\text{bond}} + U_{\text{angle}} + U_{\text{dihedral}}$$

$$U_{\text{nonbonded}} = U_{\text{van der Waals}} + U_{\text{electrostatic}}$$



MDFE

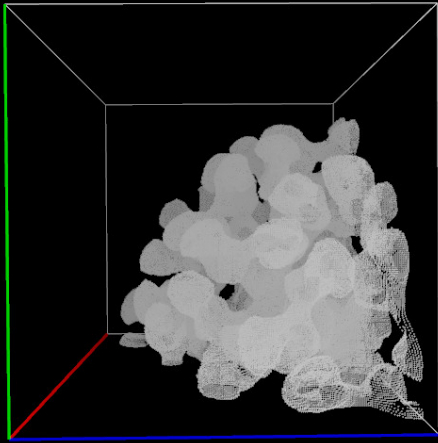
$$U_{\text{Total}} = U_{\text{bonded}} + U_{\text{nonbonded}} + U_{\text{EM}}$$



griddx
gridpdb



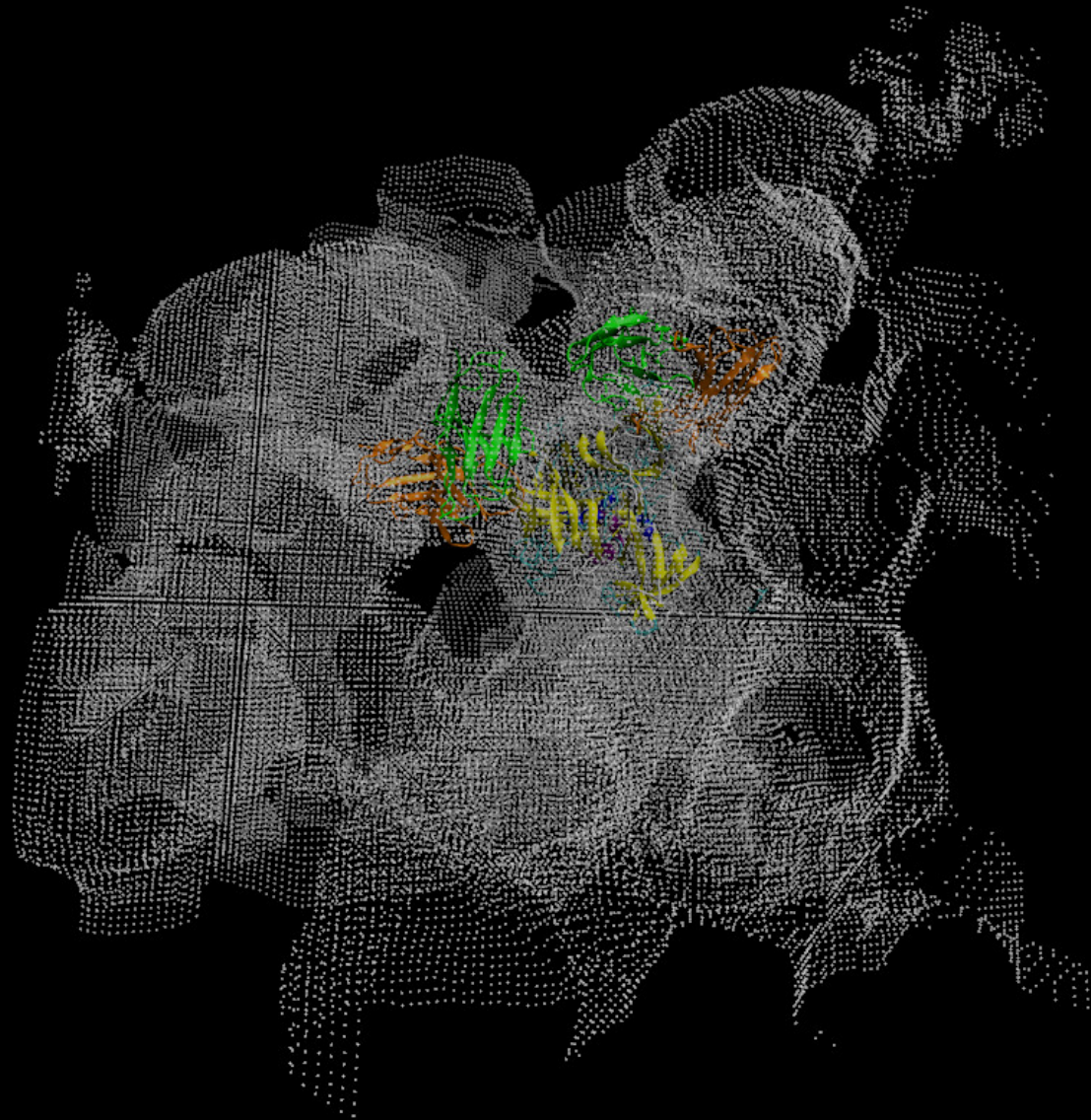
U_{em}



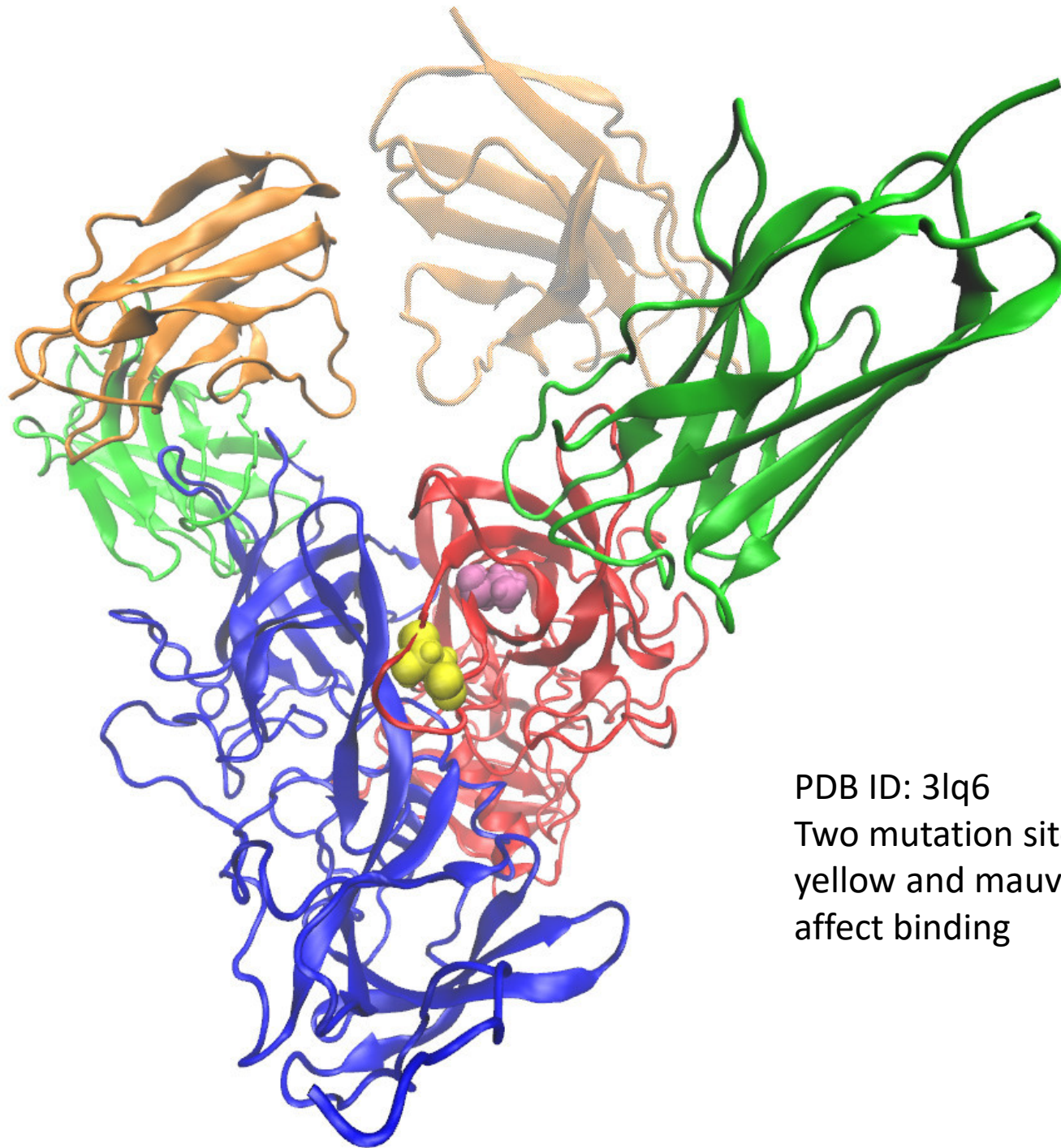
Rigid body fit with Situs

<https://situs.biomachina.org/fguide.html>

VMD ssrestraints – extra bonds to
maintain secondary structure

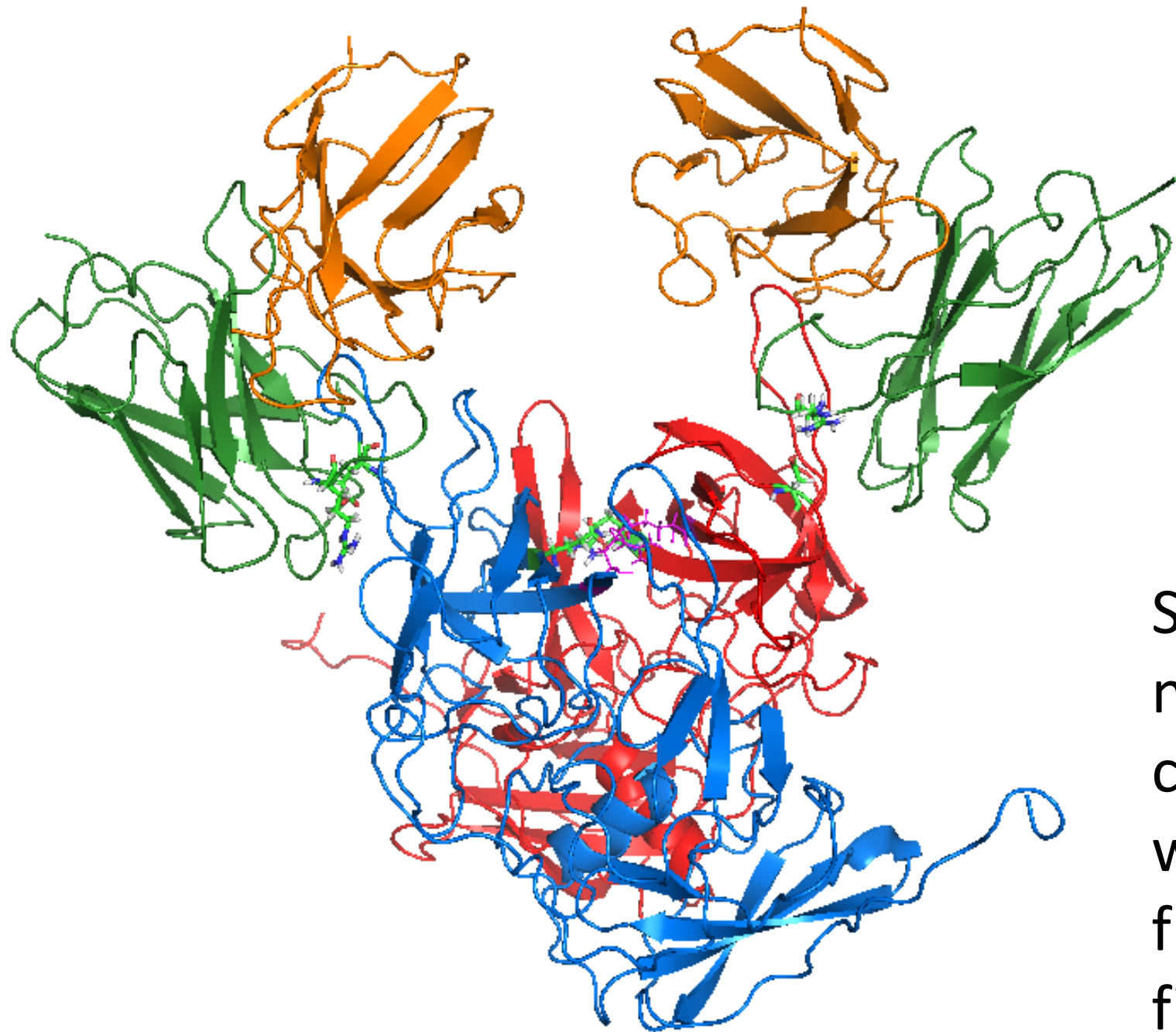


3D grid
of the
mrc file



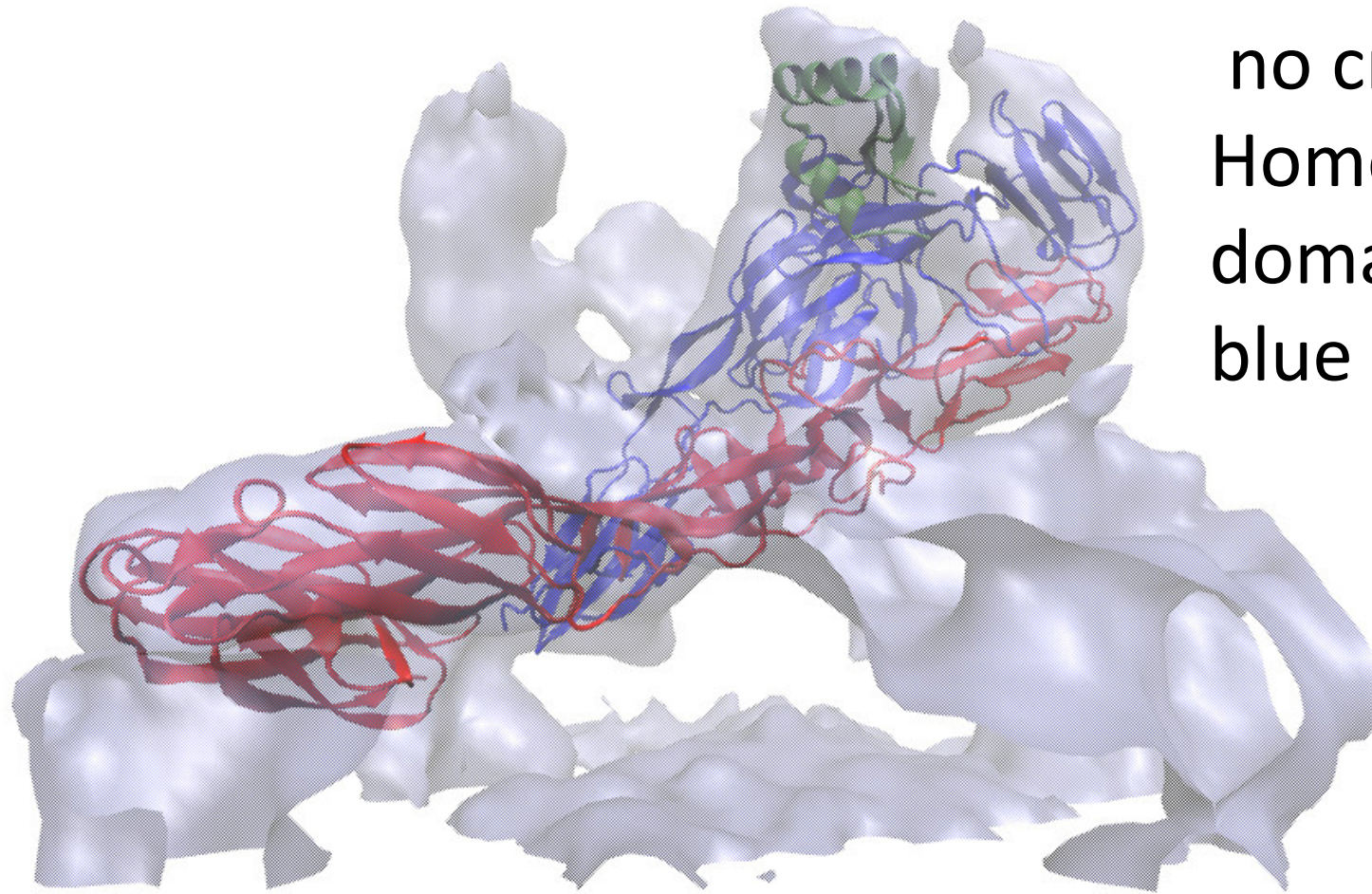
PDB ID: 3lq6

Two mutation sites shown as
yellow and mauve balls that
affect binding



Salt bridge
network
changes
with
flexible
fitting

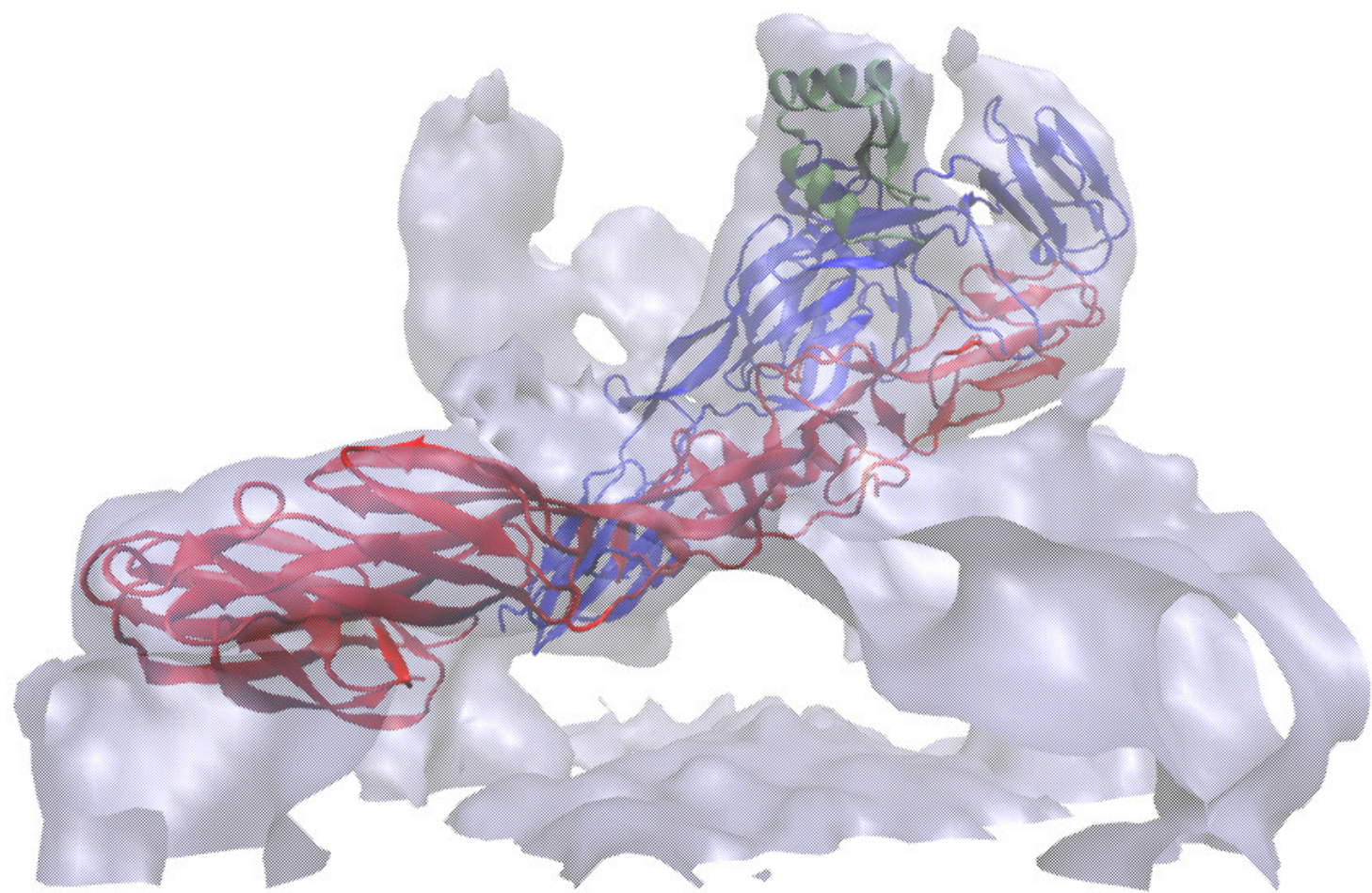
Chain A	Chain B	Cryo-EM reconstruction	MDFF Fitted R Å	V339I Fitted R Å
A:ARG 231[OD2]	B:ARG228[N]			
A:ARG 238[NE]	B:ASP 313[OD2]	3.32		
A:ARG 238[NH1]	B:ASP 313[OD2]		3.7	
A:ASP 255[OD1]	B:LYS 345[NZ]			3.97
A:ASP 313[OD1]	B:ARG 238[NH1]			
A:ASP 313[OD2]	B:ARG 238[NE]	3.14		
A:ASP 313[OD2]	B:ARG 238[NH1]	3.28		
A:ASP 313[OD2]	B:ARG 238[NH2]	3.93		
A:GLU 338[OE1]	B:ARG 396[NE]			
A:GLU 338[OE1]	B:ARG 396[NH1]	2.76		
A:GLU 338[OE1]	B:ARG 396[NH2]	3.65		
A:GLU 338[OE2]	B:ARG 396[NH1]	2.63		
A:GLU 338[OE2]	B:ARG 396[NH2]	3.12		3.44
A:GLU 338[OE2]	B:ARG 437[NH2]	2.75		
A:LYS 349[NZ]	B:ASP 440[OD2]			3.98
A:ARG 396[NH1]	B:GLU 338[OE1]	2.85		3.92
A:ARG 396[NH1]	B:GLU 338[OE2]	3.16		3.31
A:ARG 396[NH2]	B:GLU 338[OE2]	2.58		3.57
A:ARG 396[NH2]	B:GLU 338[OE1]	3.81	2.98	
A:ARG 396[NE]	B:GLU 338[OE2]			



Second system
no crystal structure
Homology model of 3
domain shown in red,
blue and green



Order of rigid body fit matters. If this structure is fitted first it could go anywhere in the map and not in the volume where it should be located

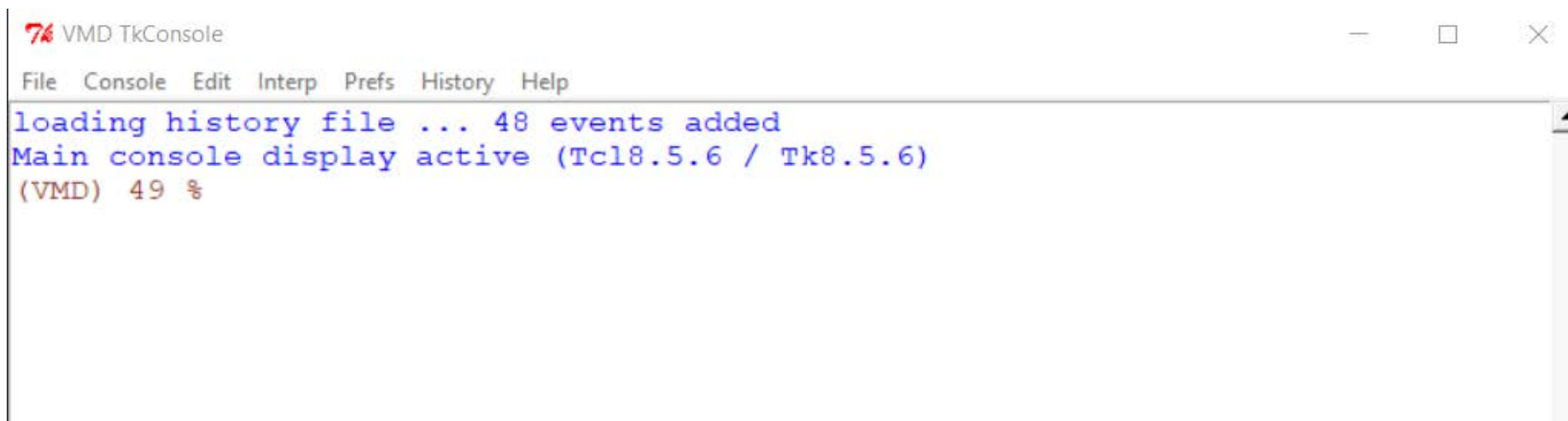


Norovirus MNV Example

RCSB PDBID: 3lq6

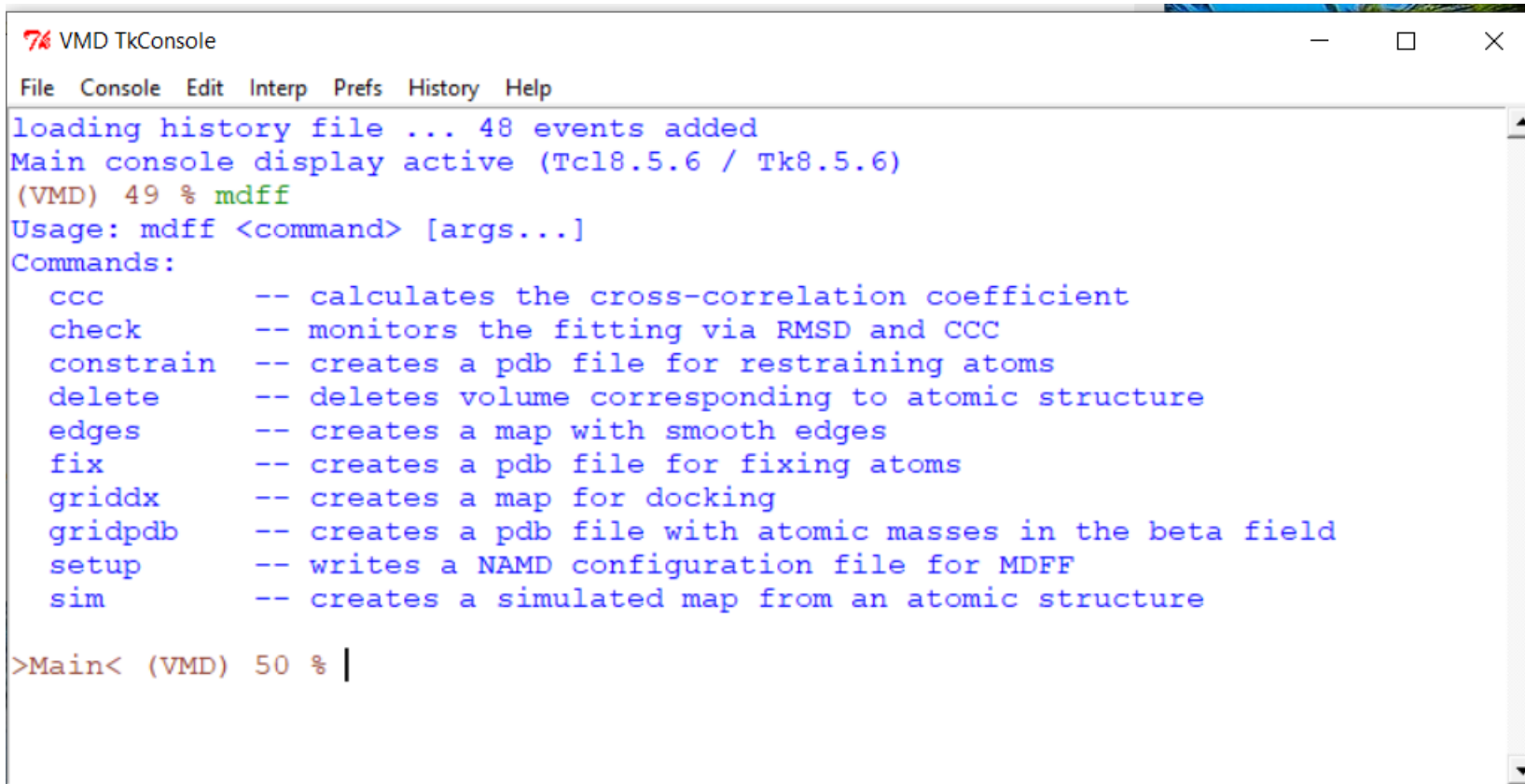
Cryo-EM Map – MNV-Fab-trimmed.mrc

Commands will be executed in VMD's Tk console



```
VMD TkConsole
File Console Edit Interp Prefs History Help
loading history file ... 48 events added
Main console display active (Tcl8.5.6 / Tk8.5.6)
(VMD) 49 %
```

VMD – use mdff griddx to generate 3D grid of map



```
VMD TkConsole
File Console Edit Interp Prefs History Help
loading history file ... 48 events added
Main console display active (Tcl8.5.6 / Tk8.5.6)
(VMD) 49 % mdff
Usage: mdff <command> [args...]
Commands:
ccc          -- calculates the cross-correlation coefficient
check       -- monitors the fitting via RMSD and CCC
constrain   -- creates a pdb file for restraining atoms
delete      -- deletes volume corresponding to atomic structure
edges       -- creates a map with smooth edges
fix         -- creates a pdb file for fixing atoms
griddx      -- creates a map for docking
gridpdb     -- creates a pdb file with atomic masses in the beta field
setup       -- writes a NAMD configuration file for MDFF
sim         -- creates a simulated map from an atomic structure

>Main< (VMD) 50 % |
```

Type the following command in the Tk console

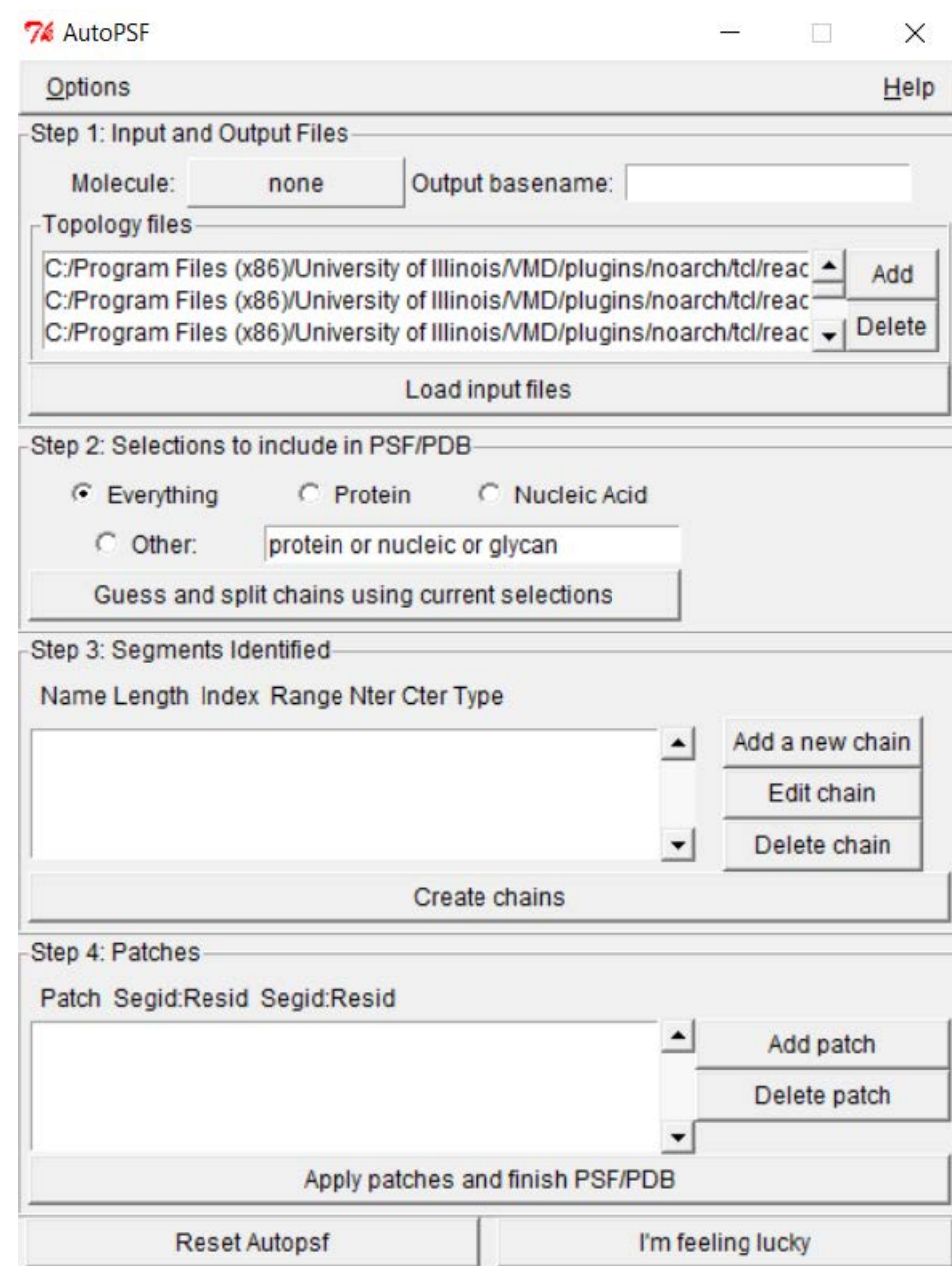
```
mdff grid dx -i MNV-Fab-trimmed.mrc -o  
MNV-Fab-trimmed.dx
```



The screenshot shows a window titled "VMD TkConsole" with a menu bar containing "File", "Console", "Edit", "Interp", "Prefs", "History", and "Help". The main text area displays the command: `>Main< (VMD) 51 % mdff grid dx -i MNV-Fab-trimmed.mrc -o MNV-Fab-trimmed.dx`. The window has standard minimize, maximize, and close buttons in the top right corner.

Use VMD to add hydrogens to the starting structure and generate a PSF file

3lq6-psf.pdb
3lq6-psf.psf



Rigid body fit the PDB to the map

If using Situs:

```
colores MNV-Fab-trimmed.mrc 3lq6-psf.pdb -res  
9.0 -powell
```

Rename the best structure

```
cp col_best_001.pdb 3lq6-psf-docked.pdb
```

Use VMD - Prepare the initial structure and files

PDB file containing the per-atom scaling factors
for U_{EM}

```
mdff gridpdb -psf 3lq6-psf.psf -pdb  
3lq6-psf-docked.pdb -o 3lq6-grid.pdb
```

Use VMD - Prepare the initial structure and files

Define secondary structure restraints

```
ssrestraints -psf 3lq6-psf.psf -pdb  
3lq6-psf-docked.pdb -o 3lq6-  
extrabonds.txt -hbonds
```

Use VMD - Prepare the initial structure and files

Retain peptide bonds to cis/trans configuration
as well as chiral centers

First – need to load the pdb and psf files

```
mol new 3lq6-psf.psf
```

```
mol addfile 3lq6-psf-docked.pdb
```

Use VMD - Prepare the initial structure and files

Use cispeptide plugin to restrain cis peptide bonds to current cis/trans config.

cispeptide restrain -o 3lq6-extrabonds-cispeptide.txt

Use VMD - Prepare the initial structure and files

Use chirality plugin to restrain chiral centers

```
chirality restrain -o 3lq6-extrabonds-chirality.txt
```

Use VMD - Prepare the initial structure and files

Generate NAMD mdff configuration file. For this simple example only 5000 steps will be used and a grid scale of 0.3


```
mdff setup -o 3lq6 -psf 3lq6-psf.psf  
-pdb 3lq6-psf-docked.pdb  
-griddx MNV-Fab-trimmed.dx  
-gridpdb 3lq6-grid.pdb  
-extrab {3lq6-extrabonds.txt 3lq6-extrabonds-  
cispeptide.txt 3lq6-extrabonds-chirality.txt}  
-gscale 0.3 -numsteps 5000
```

Two files will be created:

3lq6-step1.namd

mdff-template.namd

Run the mdff calculation with
NAMD

```
namd2 3lq6-step1.namd >& 3lq6-step1.log
```

Use VMD to view results

The final structure will be in the file named 3lq6-step1.coor. Compare the 2 structures

```
mol new 3lq6-psf.psf
```

```
mol addfile 3lq6-step1.coor type namdbin
```

```
mol new 3lq6-psf-docked.pdb
```

TACC

